This article was downloaded by: [Renmin University of China] On: 13 October 2013, At: 10:51 Publisher: Taylor & Francis Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



# Journal of Coordination Chemistry

Publication details, including instructions for authors and subscription information: http://www.tandfonline.com/loi/gcoo20

# Synthesis and studies of selective chemosensors for anions and cations by azo-containing salicylaldimine-based receptors

Raziyeh Arabahmadi<sup>a</sup> & Saeid Amani<sup>a</sup>

<sup>a</sup> Faculty of Sciences, Department of Chemistry , Arak University , Arak , Iran

Accepted author version posted online: 27 Nov 2012. Published online: 14 Jan 2013.

To cite this article: Raziyeh Arabahmadi & Saeid Amani (2013) Synthesis and studies of selective chemosensors for anions and cations by azo-containing salicylaldimine-based receptors, Journal of Coordination Chemistry, 66:2, 218-226, DOI: <u>10.1080/00958972.2012.752818</u>

To link to this article: <u>http://dx.doi.org/10.1080/00958972.2012.752818</u>

# PLEASE SCROLL DOWN FOR ARTICLE

Taylor & Francis makes every effort to ensure the accuracy of all the information (the "Content") contained in the publications on our platform. However, Taylor & Francis, our agents, and our licensors make no representations or warranties whatsoever as to the accuracy, completeness, or suitability for any purpose of the Content. Any opinions and views expressed in this publication are the opinions and views of the authors, and are not the views of or endorsed by Taylor & Francis. The accuracy of the Content should not be relied upon and should be independently verified with primary sources of information. Taylor and Francis shall not be liable for any losses, actions, claims, proceedings, demands, costs, expenses, damages, and other liabilities whatsoever or howsoever caused arising directly or indirectly in connection with, in relation to or arising out of the use of the Content.

This article may be used for research, teaching, and private study purposes. Any substantial or systematic reproduction, redistribution, reselling, loan, sub-licensing, systematic supply, or distribution in any form to anyone is expressly forbidden. Terms & Conditions of access and use can be found at <u>http://www.tandfonline.com/page/terms-and-conditions</u>



# Synthesis and studies of selective chemosensors for anions and cations by azo-containing salicylaldimine-based receptors

# RAZIYEH ARABAHMADI and SAEID AMANI\*

Faculty of Sciences, Department of Chemistry, Arak University, Arak, Iran

(Received 8 July 2012; in final form 11 October 2012)

Three new chromogenic receptors containing electron-withdrawing groups appended to the azophenol moiety were synthesized, characterized, and their chromogenic behavior toward various anions and cations investigated. These chemosensors show visual changes towards biologically relevant anions like  $F^-$  and  $OH^-$ , and also towards various cations such as  $Cu^{2+}$ ,  $Co^{2+}$ ,  $Ni^{2+}$ ,  $Mn^{2+}$ , and  $Zn^{2+}$ . Yet, other anions such as  $Cl^-$ ,  $Br^-$ , and  $NO_3^-$  could not cause any color change. The sensing action was further confirmed by UV–Vis titration. <sup>1</sup>H-NMR experiments were carried out to explore the nature of interaction between receptors with  $F^-$  and  $OH^-$ . The binding constants ( $K_a$ ) of the host–guest complexes formed were determined.

Keywords: Chemosensor; Transition metal azo complexes; Receptors

# 1. Introduction

Considerable attention has been focused on recognition and sensing of anions and cations since these play important roles in biology, industry, and catalysis [1]. Generally, pyrrole, urea/thiourea, amine, or phenol act as binding sites for anions [2–4] and cations [5–8]. In particular, sensing of fluoride has attracted growing attention due to unique properties compared to its congeners as a result of high electronegativity, smaller size, and high basicity. Among anions, fluoride is of particular interest owing to its established role in treatment of osteoporosis and analysis of drinking water [9, 10]. Currently available methods for detection of fluoride include spectrophotometry, electrochemistry, and ion chromatography [11–13]. From the pharmacological and environmental importance of fluoride, there is need to develop selective and sensitive methods for fluoride detection in the environment that is not easily served by conventional ion selective electrodes. In this context, colorimetric chemosensors are of particular interest due to their simplicity. Colorimetric chemosensors allow "naked-eye" detection of anions without need to resort to spectroscopic instrumentation [14]. Most colorimetric chemosensors only respond to anions or cations separately [15–18]. Development of chemosensors for detection of transition metal cations is active due to potential applications as diagnostic tools in medical, physiological, and environmental applications [19]; transition metal ions are present as essential elements in biological systems [20].

<sup>\*</sup>Corresponding author. Email: s-amani@araku.ac.ir

We report for the first time the colorimetric studies of cation and anion sensing by chromogenic receptors 2a-2c possessing a phenolic OH, which binds F<sup>-</sup> and OH<sup>-</sup> via H–bond interaction or deprotonation, with an electron-withdrawing nitro which acts as a chromogenic signaling unit. The nature of these Schiff bases was altered by the position of nitro in the receptors, which was able to tune anion recognition selectivity. These receptors are selective sensors for Cu<sup>2+</sup>, Co<sup>2+</sup>, Ni<sup>2+</sup>, Mn<sup>2+</sup>, and Zn<sup>2+</sup> in DMSO (Scheme 1).



# 2. Experimental

# 2.1. Physical measurements

C, H, and N analyses were undertaken using an elemental analysis system Gmb H Vario EL III. Electronic spectra of receptors in DMSO were recorded on a Perkin-Elmer Lambda 15 spectrophotometer. FT-IR spectra of the compounds as KBr disks were obtained from 4000 to 300 cm<sup>-1</sup> with a Galaxy series FT-IR 5000 spectrophotometer. Melting points of all newly prepared compounds were determined on an Electrothermal 9200 apparatus. <sup>1</sup>H-NMR spectroscopy was performed using a Bruker 300 MHz spectrometer.

### 2.2. Materials

All anions as tetrabutylammonium salts were purchased from Fluka and used without purification. All solvents were of reagent grade quality, purchased from Merck Chemical Company and used as received. The  $Cu^{2+}$ ,  $Co^{2+}$ ,  $Ni^{2+}$ ,  $Mn^{2+}$ , and  $Zn^{2+}$  were added as chloride solutions in DMSO.

# 2.3. Synthesis

Azo dyes (1a-1c) were synthesized according to the well-known literature procedure [21].

**2.3.1.** Synthesis of the Schiff bases. A mixture of 0.02 M of 2-(2-aminoethoxy) ethanol and 0.02 M of the related azo dye (1a–1c) was dissolved in 50 mL absolute ethanol with a few drops of glacial acetic acid as a catalyst. The resulting mixture was allowed to stir magnetically under reflux for 1–2 h. The product was filtered under vacuum and washed with a small amount of heated ethanol.

Synthesis of 1-{3-[(1-ethoxy-2-hydroxyethyl-imino)methyl]-4-hydroxyphenylazo}-4-nitro benzene (2a). Brownish red, yield: 63%, m.p.: 174 °C. Anal. Calcd for  $C_{17}H_{18}N_4O_5 \cdot 0.5$ - $CH_3OH$ : C, 56.17; H, 5.34; N, 14.96. Found: C, 56.70; H, 6.17; N, 15.22%. FT-IR (KBr,  $cm^{-1}$ ): 3110–3650 (–OH group), 3105 (C–H, aromatic), 2874 (C–H, aliphatic), 1651 (C=N), 1614 (C=C, aromatic), 1421 (N=N), 1271 (C–O, phenolic) cm<sup>-1</sup>. UV–Vis (DMSO, nm):  $\lambda_{max}$ =260 (14,150), 407 (16,125), 449 (16,775) nm. <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$ =10–11 (1H, s), 8.46 (1H, s), 8.39 (2H, d, J=9.0 Hz), 8.03 (2H, dd, J=9 Hz), 7.99 (2H, d, J=7 Hz), 7.97 (1H, d, J=2.4 Hz) 7.0 (1H, d, J=9.03 Hz), 3.87 (4H, m, J=3.6 Hz), 3.77 (2H, t, J=3.3 Hz), 3.64 (2H, t, J=4.02 Hz) ppm.

Synthesis of  $1-\{3-[(1-ethoxy-2-hydroxethyl-imino)methyl]-4-hydroxyphenylazo\}-2-chloro 4-nitrobenzene (2b). Dark brown, yield: 46%, m.p.:185 °C. Anal. Calcd for C<sub>17</sub>H<sub>17</sub>N<sub>4</sub>O<sub>5</sub>Cl·H<sub>2</sub>O: C, 49.71; H, 4.62; N, 13.63. Found: C, 50.40; H, 4.60; N, 13.87%. FT-IR (KBr, cm<sup>-1</sup>): 3120–3700 (–OH group), 3115 (C–H, aromatic), 2889 (C–H, aliphatic), 1651 (C=N), 1614 (C=C, aromatic), 1408 (N=N), 1269 (C–O, phenolic) cm<sup>-1</sup>. UV–Vis (DMSO, nm): <math>\lambda_{max} = 260$  (13,525), 407 (14,575), 469 (17,375) nm. <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta = 10.2$  (1H, s), 8.44 (1H, s), 8.42 (1H, d, J = 2.2 Hz), 8.1 (1H, dd, J = 8.7, 2.2 Hz), 8.0 (1H, dd, J = 9.0 Hz), 7.9 (1H, d, J = 2.0 Hz), 7.7 (1H, d, J = 8.8 Hz), 7.0 (1H, d, J = 9.0 Hz), 3.86 (4H, m, J = 4.8 Hz), 3.78 (2H, t, J = 4.5 Hz), 3.65 (2H, t, J = 4.2 Hz) ppm.

Synthesis of 1-{3-[(1-ethoxy-2-hydroxyethyl-imino)methyl]-4-hydroxyphenylazo}-4-chloro-3nitrobenzene (2c). Light brown, yield: 32%, m.p.: 223 °C. Anal. Calcd for C<sub>17</sub>H<sub>17</sub>N<sub>4</sub>O<sub>5</sub>Cl·-CH<sub>3</sub>OH: C, 50.90; H, 4.94; N, 13.18. Found: C, 50.40; H, 3.69; N, 13.89%. FT-IR (KBr, cm<sup>-1</sup>): 3300–3700 (–OH group), 3200 (C–H, aromatic), 2918 (C–H, aliphatic), 1647 (C=N), 1616 (C=C, aromatic), 1406(N=N), 1288 (C–O, phenolic) cm<sup>-1</sup>. UV–Vis (DMSO, nm):  $\lambda_{max}$ =262 (17,600), 387 (19,575), 430 (18,875) nm. <sup>1</sup>H-NMR (CDCl<sub>3</sub>):  $\delta$ =10 (1H, s),8.47 (1H, s), 8.36 (1H, d, *J*=2.2 Hz), 8.06 (1H, dd, *J*=17.4, 2.2 Hz), 8.02 (1H, d, *J*=2.3 Hz), 7.9 (1H, d, *J*=2.3 Hz), 7.7 (1H, d, *J*=3.6 Hz), 7.0 (1H, d, *J*=9 Hz), 3.8 (4H, m, *J*=3.3 Hz), 3.7 (2H, t, *J*=4.5 Hz), 3.6 (2H, t, 3.9 Hz) ppm.

## 3. Results and discussion

#### 3.1. Anion sensing

Recognition properties of 2a-2c towards different anions were studied by methods such as naked-eye examination, the <sup>1</sup>H-NMR titration, and UV-visible titration.

**3.1.1. Colorimetric investigations.** In the naked-eye experiments, receptors 2a-2c showed dramatic color changes in the presence of two equivalents of tetrabutylammonium fluoride ( $5 \times 10^{-4}$  M in DMSO). Receptor 2a displays noticeable color change from light pale orange to violet and 2b and 2c gave dramatic color changes from yellow to blue and orange, respectively. In addition, 2a-2c could sense OH<sup>-</sup> (see Supplementary Material, figure S The receptors 2a-2c did not show color change for Cl<sup>-</sup>, Br<sup>-</sup>, and NO<sub>3</sub><sup>-</sup>. The receptors bound anions through hydrogen bonds to form stable complexes and the electron density in the supramolecular system was increased to enhance the charge-transfer interactions between the electron-rich and electron-deficient moieties, resulting in a visible color change [22]. Restoration of the original spectrum of the sensor system from this receptor F<sup>-</sup> adduct upon addition of a trace amount of water/methanol suggests that complexations between F<sup>-</sup> and OH<sup>-</sup> with 2a-2c are reversible in nature [23].

Electronic spectra of the receptors showed three transitions in DMSO. The first band at 260 nm was assigned to excitation of the  $\pi$ -electrons of the aromatic system. The second band observed at 387 and 407 nm could be due to the transition between the  $\pi$ -orbital localized on the central bond of the azomethine group (CH=N). The third band located at 449, 469, and 430 nm is due to an intramolecular charge transfer (CT) transition within the whole molecule. This important band in salicylaldimine compounds indicates the presence of a strong intramolecular hydrogen bond between hydroxyl and azomethine nitrogen that causes planarity and facilitates CT bands which are more sensitive to solvent changes than bands resulting from local transitions [24, 25].

**3.1.2.** <sup>1</sup>H-NMR spectral studies. We investigated the interaction of receptors with anions through <sup>1</sup>H-NMR studies. In <sup>1</sup>H-NMR spectra of the receptors, singlet resonances in the 10–10.3 ppm region can be attributed to protons of phenol. All three receptors exhibit singlets at 8.36–8.46 ppm, which were attributed to imine protons. <sup>1</sup>H-NMR spectra were recorded by treating the receptor with F<sup>-</sup> and OH<sup>-</sup>. The peak at 10.3 ppm corresponding to proton of phenolic OH disappeared on addition of one equivalent of F<sup>-</sup> or OH<sup>-</sup>. This demonstrates the interaction of anion through the phenolic OH is more effective than interaction through the alcoholic OH groups present in the receptor. Deprotonation of the receptor can occur and we observed such deprotonation with formation of HF<sub>2</sub><sup>-</sup> in <sup>1</sup>H-NMR spectra of **2b** as a new signal at 16 ppm [figure 1(a)]. A similar case can be observed for addition of OH<sup>-</sup> as shown in figure 1(b) [26]. The interesting part of the NMR studies is in figure 1(B) trace C. There is a large displacement in the signals. Further <sup>1</sup>H-NMR studies are in progress to understand this behavior.

**3.1.3. UV–Vis spectroscopic studies.** The sensing behavior of 2a-2c towards F<sup>-</sup>, Cl<sup>-</sup>, Br<sup>-</sup>, NO<sub>3</sub><sup>-</sup>, and OH<sup>-</sup> was also investigated by UV–Vis spectroscopy. UV–Vis titration was carried out by incremental addition of tetrabutylammonium salts to 2a-2c and the spectra are shown in figure 2. When increasing the concentration of F<sup>-</sup> in 2a-2c, a new red shifted absorption at 565, 583, and 503 nm was gradually enhanced, while the intensity of absorptions at 449, 469, and 387 nm decreased correspondingly. Clear isosbestic points were observed at 492, 524, and 403 nm for 2a-2c, respectively. The appearance of a single isosbestic point indicates the presence of only two species, neutral host and its anion in solution. Figure 2(d) shows the gradual decrease and



Figure 1. (a) Partial <sup>1</sup>H-NMR (300 MHz) spectra of **2b** in CDCl<sub>3</sub>, (A) in the absence, (B) presence of 1.0 equiv., and (C) presence of 3.0 equiv. of TBAF. (b) Partial <sup>1</sup>H-NMR (300 MHz) spectra of **2b** in CDCl<sub>3</sub>, (A) in the absence, (B) presence of 1.0 equiv., and (C) presence of 3.0 equiv. of TBAOH.

increase in the concentration of these two species with increase in equivalents of tetrabutylammonium fluoride, respectively. Similar changes could be observed during the addition of  $OH^-$  (see Supplementary Material, figure S2). The negative charge brought about by anion-induced deprotonation increases the dipole moment and stabilizes the excited state, causing a red shift of **2a–2c**. These results demonstrate that complex-formation of **2a–2e** with  $F^-$  and  $OH^-$  takes place via hydrogen bonding electrostatic interactions (table 1). No change was observed with  $CI^-$ ,  $Br^-$ , and  $NO_3^-$  in DMSO [figure 2(e)] (see Supplementary Material, figures S3–S5).

The chromogenic diazo unit is introduced at the 5-position of salicylaldehyde to achieve direct resonance between them [27–33]. As shown in figure 3, the electron-withdrawing nitro group at the end increases not only the degree of  $\pi$ -conjugation but also the hydrogen bond donor ability of the phenolic OH. The strong hydrogen bonding to or deprotonation/ protonation of the phenolic moiety might modulate the electronic properties of the chromophore and give rise to large color changes [25].



Figure 2. (a-c) UV-Vis spectra of 2a-2c in the course of titration of DMSO solutions of 2a-2c ( $4 \times 10^{-5}$  M) with a standard solution of TBAF ( $c \approx 5 \times 10^{-4}$  M) at 25 °C; inset shows the visual color change on addition of F<sup>-</sup>. (d) Plot of 2a absorption vs. concentration of F<sup>-</sup> showing decrease and increase in absorption of the 449 nm and 565 nm bands, respectively. (e) UV-Vis absorption spectra of 2a upon addition of a particular anion salt.

Table 1. Data obtained from the UV–Vis spectra upon titration of 2a-2c with n-Bu<sub>4</sub>N<sup>+</sup>F<sup>-</sup> in DMSO.

	Receptor, $\lambda_{max}$ (nm)	Complex, $\lambda_{\max}$ (nm)	Bathochromic shift, $\Delta \lambda_{\max}$ (nm)	Isosbestic point (nm)	$K_{\rm a}  ({ m M}^{-1})$	Detection limit (M)
2a	449	565	114	492	$8.616 \times 10^3$	$1.64 \times 10^{-6}$
2b	469	583	117	524	$3.7 \times 10^{3}$	$2.19 \times 10^{-6}$
2c	387	503	98	403	$1.262 \times 10^{3}$	$2.55 \times 10^{-6}$

### 3.2. Calculation of binding constant

**3.2.1.** UV–Vis titration. The binding constants of **2a–2c** were calculated from the fluoride-induced absorption changes using equation (1) [34],

$$\frac{b}{\Delta A} = \frac{1}{S_{t}K_{a}\Delta\varepsilon} \times \frac{1}{[L]} + \frac{1}{S_{t}\Delta\varepsilon}$$
(1)

where  $\Delta A$  is the change in absorbance, *b* is path length, *S*<sub>t</sub> is total concentration of substrate, *K*<sub>a</sub> is association constant, and  $\Delta \varepsilon$  is the change in molar absorption coefficient. A plot can be made of  $1/\Delta A$  as a function of 1/[L]. The binding constant can be obtained by the ratio of intercept and slope (see Supplementary Material, figures S6 and S7). The detection limits of the sensors toward F<sup>-</sup> and OH<sup>-</sup> were obtained from UV–Vis titration. The result can also be calculated from the fluoride quantities added, which made the initial changes in absorbance of the sensors during UV–Vis titration [35]. The binding constants and detection limits are summarized in table 1.

## 3.3. Cation sensing

We investigated the recognition ability of 2a-2c ( $2 \times 10^{-5}$  M in DMSO) by naked-eye colorimetric experiments for Cu<sup>2+</sup> ( $2.5 \times 10^{-3}$  M), Co<sup>2+</sup> ( $8 \times 10^{-3}$  M), Ni<sup>2+</sup> ( $3 \times 10^{-2}$  M), Mn<sup>2+</sup> ( $6 \times 10^{-2}$  M), and Zn<sup>2+</sup> ( $6 \times 10^{-2}$  M) (for color change of 2a-2c, see Supplementary Material, figures S8–S10).

The sensing behavior of 2a-2c towards  $Cu^{2+}$ ,  $Co^{2+}$ ,  $Ni^{2+}$ ,  $Mn^{2+}$ , and  $Zn^{2+}$  was monitored by the spectrophotometric method in DMSO. A solution of 2a-2c was titrated with increasing volume of concentrated solution of a given cation. On incremental addition of  $Cu^{2+}$  to 2a, the peak at 260 nm gradually increased with a red shift and a new band arose at 885 nm. On further addition of  $Co^{2+}$  to 2a a new band was formed at 623, 685 nm. On gradual addition of  $Ni^{2+}$  to 2a, a new absorption band at 805 nm appeared. On successive addition of  $Mn^{2+}$  and  $Zn^{2+}$ , the spectrum of 2a shifted from 460 to 470 and 447 nm. Similar changes were observed for 2b and 2c. The insert plot in the UV–Vis spectrum shows the variation of absorbance at a particular wavelength vs. equivalents of the corresponding ion added (for titration spectrum, see Supplementary Material, figures S11–S13).



Figure 3. Resonance structures of phenolic OH and chromogenic diazo scaffold.

## 3.4. Sensing in the presence of competing ions

Since 2a-2c sense both transition metal ions and F<sup>-</sup> and OH<sup>-</sup>, we examined the sensing ability in the presence of competing ions. These experiments were carried out in two different manners. First, the receptors were treated with two equivalents of anions and to this, successive addition of cations (20-200 µl) was done; in another set of experiments. the receptors were first treated with two equivalents of cations and to that  $20-200 \,\mu$ l of anion was added incrementally. For this dual-sensing experiment, Cu2+ and F- were chosen. The spectral changes are shown in figures S14-S16 (see Supplementary Material). When 2a was treated with two equivalents of  $Cu^{2+}$ , a new band was observed at 805 nm which might be due to the d-d band of  $2a \cdot Cu^{2+}$  complex. Addition of F<sup>-</sup> (0.5-2 equiv.) ions to the above  $(2a \cdot Cu^{2+})$  solution leads to reduction in intensity of the band at 805 nm, possibly due to formation of a Cu:F ion pair ion in solution. In the second experiment, the titration was carried out in the presence of two equivalents of  $F^-$  to which  $Cu^{2+}$  was added incrementally. When **2a** was added to two equivalents of F<sup>-</sup>, the intensity of the band at 449 nm decreased and a new band at 565 nm indicated formation of  $2a \cdot F^-$  complex. Upon successive addition of  $Cu^{2+}$  the band at 565 nm gradually decreased, suggesting sequestering of the  $F^-$  to form an ion pair with  $Cu^{2+}$  in solution. Intensity of the d-d band increased slightly, possibly due to interaction between 2a and  $Cu^{2+}$  ions.

In another experiment, **2a** was titrated with a mixture of  $F^-$  and  $Cu^{2+}$  (using 15, 30, 45 and 60 equiv.). When the mixture of  $F^-$  with  $Cu^{2+}$  was introduced to the DMSO solution of **2a**, the intensity of the band at 260 nm increased and enhancement of the d–d band occurred, due to formation of a  $Cu^{2+}$  complex with **2a**. The formation of the band at 565 nm was not observed in this case because  $F^-$  in solution engaged the metal ions in ion-pair formation rather than in complexation with these receptors. Similar changes were observed for **2b** and **2c** (see Supplementary Material, figures S17–S22). We repeated similar experiments with  $F^-$  and  $Co^{2+}$  showing the receptors **2a–2c** can be utilized for the detection of  $Co^{2+}$  in the presence of competing  $F^-$  (mixture of  $F^-$  and  $Co^{2+}$ ) in DMSO by change in absorption spectra.

#### 4. Conclusion

Receptors 2a-2c are prepared via diazo coupling and Schiff base condensation, and possess the ability to sense both anions and cations colorimetrically. These Schiff base receptors form metal complexes and sense anions through hydrogen bonding interaction through the phenolic OH. In 2a, the presence of the electron-withdrawing NO<sub>2</sub> enhances hydrogen bonding and exhibits significant difference in anion binding by providing colorimetric spectral responses. These receptors sense Cu<sup>2+</sup> selectively in the presence of F<sup>-</sup>, confirmed by the absorbance spectra of the competing studies.

### Acknowledgement

The authors would like to thank the Research Council of Arak University for financial support of this research.

Downloaded by [Renmin University of China] at 10:51 13 October 2013

### References

- [1] R. Martinez-Manez, F. Sancenon. Chem. Rev., 103, 4419 (2003).
- [2] J.L. Sessler, H. Maeda, T. Mizuno, V.M. Lynch, H. Furuta. Chem. Commun., 862, (2002).
- [3] H.J. Kim, J.H. Lee, T.H. Kim, W.S. Lyoo, D.W. Kim, T.S. Lee. J. Polym. Sci. Part A: Polym. Chem., 45, 1546 (2007).
- [4] M. Yu, H. Lin, G. Zhao, H. Lin. J. Mol. Recognit., 20, 69 (2007).
- [5] P. Kar, M. Suresh, D.K. Kumar, D.A. Jose, B. Ganguly, A. Das. Polyhedron, 26, 1317 (2007).
- [6] J.S. Kim, O.J. Shon, J.K. Lee, S.H. Lee, J.Y. Kim, S. Lee. J. Org. Chem., 67, 1372 (2002).
- [7] Y. Kubo. J. Inclusion Phenom. Mol. Recognit. Chem., 32, 235 (1998).
- [8] T. Ghosh, B.G. Maiya, A. Samanta. Dalton Trans., 795 (2006).
- [9] K.L. Krick. Biochemistry of the Halogens and Inorganic Halides. p. 58, Plenum, New York, NY (1991).
- [10] S. Ayoob, A.K. Gupta. Crit. Rev. Env. Sci. Technol., 36, 433 (2006).
- [11] Y.F. Cheng, Z.Q. Liu, M. Shi, Q. Zhao, F.Y. Li, C.H. Huang. Chin. J. Chem., 25, 616 (2007).
- [12] S. Sathish, G. Narayan, N. Rao, C. Janardhana. J. Fluoresc., 17, 1 (2007).
- [13] S. Jeyakumar, V.V. Raut, K.L. Ramakumar. Talanta, 76, 1246 (2008).
- [14] J. Xu, K. Liu, D. Di, S. Shao, Y. Guo. Inorg. Chem. Commun., 10, 681 (2007).
- [15] T. Gunnlaugsson, P.E. Kruger, P. Jensen, J. Tierney, H.D.P. Ali, G.M. Hussey. J. Org. Chem., 70, 10875 (2005).
- [16] B. Liu, H. Tian. Chem. Lett., 34, 686 (2005).
- [17] J.V. Ros-Lis, M.D. Marcos, R. Martinez-Manez, K. Rurack, J. Soto. Angew. Chem. Int. Ed., 44, 4405 (2005).
- [18] S.J. Lee, S.S. Lee, I.Y. Jeong, J.Y. Lee, J.H. Jung. Tetrahedron Lett., 48, 393 (2007).
- [19] U.E. Spichiger-Keller (Ed.). Chemical Sensors and Biosensors for Medical and Biological Applications, Wiley-VCH, Weinheim (1998).
- [20] D. Fu, D. Yuan. Spectrochim. Acta Part A Mol. Biomol. Spectrosc., 66, 434 (2007).
- [21] R. Arabahmadi, S. Amani. Molecules, 17, 6434 (2012).
- [22] D.H. Lee, H.Y. Lee, K.H. Lee, J.I. Hong. Chem. Commun., 13, 1188 (2001).
- [23] T. Ghosh, B.G. Maiya, M.W. Wong. J. Phys. Chem. A, 108, 11249 (2004).
- [24] A.A. Gabr. Spectrochim. Acta Part A Mol. Biomol. Spectrosc., 46, 1751 (1990).
- [25] H.H. Hammud, A. Ghannoum, M.S. Masoud. Spectrochim. Acta Part A Mol. Biomol. Spectrosc., 63, 255 (2006).
- [26] K. Ghosh, G. Masanta. Tetrahedron Lett., 47, 2365 (2006).
- [27] R. Gu, S. Depraetere, J. Kotek, J. Budka, E. Wagner-Wysiecka, W. Dehaen. Org. Biomol. Chem., 3, 2921 (2005).
- [28] R. Nishiyabu, M.A. Palacios, W. Dehaen, P. Anzenbacher, Jr. J. Am. Chem. Soc., 128, 11496 (2006).
- [29] J.V. Ros-Lis, R. Martunez-Mainez, F. Sancenoin, J. Soto, K. Rurack, H. Wei Hoff. Eur. J. Org. Chem., 244, 2449 (2007).
- [30] T.D. Thangadurai, N.J. Singh, I.C. Hwang, J.W. Lee, R.P. Chandran, K.S. Kim. J. Org. Chem., 72, 5461 (2007).
- [31] N. Gimeno, X. Li, J.R. Durrant, R. Vilar. Chem. Eur. J., 14, 3006 (2008).
- [32] H. Lee, S.S. Lee. Org. Lett., 11, 1393 (2009).
- [33] W.L. Daniel, M.S. Han, J.S. Lee, C.A. Mirkin. J. Am. Chem. Soc., 131, 6362 (2009).
- [34] J. Xiong, L. Sun, Y. Liao, G.-N. Li, J.L. Zuo, X.Z. You. Tetrahedron Lett., 52, 6157 (2011).
- [35] J.-Q. Li, T.-B. Wei, Q. Lin, P. Li, Y.-M. Zhang. Spectrochim. Acta Part A Mol. Biomol. Spectrosc., 83, 187 (2011).